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SYNTHETIC STUDY ON APLYSIATOXINS, HIGHLY INFLAMMATORY AGENTS AND TUMOR PROMOTERS SYNTHESIS OF THE THREE OPTICALLY ACTIVE SEGMENTS

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The main segments in debromoaplysiatoxin and related aplysiatoxins have been efficiently synthesized from D-glucose and L- and D-tartaric acids.

Both aplysiatoxin and debromoaplysiatoxin, highly inflammatory agents, were first isolated from the digestive gland of the sea hare <u>Stylocheilus longicauda</u>.¹ Since then, a number of related toxins have been isolated from blue-green algae, <u>Lyngbya majuscula</u>, <u>Schizothrix calcicola</u>, <u>Oscillatoria nigroviridis</u> and others,^{2,3} and their stereostructures including the absolute configuration have also been elucidated on the basis of optical and ¹H NMR spectral data together with an X-ray crystallographic analysis of dibromoaplysiatoxin.³ The challenging structures of these natural products have prompted many synthetic chemists to investigate the total synthesis.⁴ From structural and physiological points of view, we are interested in debromoaplysiatoxin (1),⁵ which shows some antineoplastic activity⁶ and has also been found to be one of the powerful tumor promoters.⁷



In retrosynthetic analysis, 1 seems to be derived from the corresponding diketone (2), the structure of which is mainly divided into three segments [A, B and C], as seen in 2. In the present paper, we describe synthetic results of these segments starting from D-glucose, L-tartaric acids and D-tartaric acid. These synthesized segments might also be applicable for the syntheses of other aplysiatoxins.

Synthesis of segment A.

The readily available tosylate $(3)^8$ was converted to a chloro benzyl ether $(4)^9$ in three steps. Acid hydrolysis of 4 followed by NaBH₄ reduction afforded an open-chain sugar, which was again protected with 2,2-dimethoxypropane to give 5 and 6^9 in 50 and 46% yields, respec-

tively. Acidic isomerization of **6** afforded further desired ketal (**5**) in 91% yield, which was treated with NaOMe to give the corresponding epoxide (**7**)⁹ in quantitative yield. On methylation of **7** with Me₂CuLi, an alcohol (**8**)⁹ was produced as a main product (regioselectivity: ca. 11:1) in 81% yield. The alcohol (**8**) was further derivatized in two steps to another alcohol (**9**).⁹ Chain elongation of **9** was performed with Swern oxidation followed by Wittig reaction using the corresponding phosphorane and then catalytic hydrogenation to give a ketone (**10**)⁹ in moderate yield, which did not accompany such by-products as an epimer at the C5 carbon. After evaluation of asymmetric reductions of the carbonyl group at C₈ in **10**. Brown reduction using diisopinocampheylchloroborane¹⁰ followed by methylation with MeI-NaH promised a successful result, in which only one desired methyl ether (**11**)⁹ was obtained in ca. 100% ee¹¹ and 74% overall yield. This ether (**11**) was readily converted to the corresponding phonol (**12**) in quantitative yield, the CD curve ([θ]₂₆₀ +404° (EtOH)) of which was quite similar to that of debromoaplysiatoxin (**1**).³ The compound (**11**) was then subjected to a four step



a. i) NaOMe/BnOH-THF, room temp, overnight (95%); ii) MsCi, pyr. (100%); iii) ⁿBu₄NCi/ PhH, reflux temp, overnight (94%). b. i) IR-120 (H^{*}) dioxane-H₂O, reflux temp, 15 h (100%); ii) NaBH₄/ aq. MeOH (88%); iii) 2,2-dimethoxypropane, TsOH/acetone (5: 50%, **6**: 46%). c. TsOH (cat.)/ acetone, room temp, overnight (5: 53%, **6**: 42%). d. NaOMe/MeOH, $0 \rightarrow 40^{\circ}$ C, 2 h (100%). e. Me₂CuLi/Et₂O, -30 \rightarrow 0°C, 3 h (81%). f. i) TBDMSOTf, Et₃N/CH₂Cl₂, -10 \rightarrow 0°C, 30 min (100%); ii) H₂, Pd-C/MeOH, overnight (100%). g. i) Swern oxi.; ii) Ph₃P=CHCOC₆H₄OBn/PhMe, reflux temp, 7.5 h (71% in two steps); iii) H₂, Pd-C/EtOAc, room temp, 20 min (85%). h. i) Ipc₂BCi/THF, -25°C, overnight; MeI, NaH/THF (74% in two steps). i. H₂, Pd-C/MeOH (quantitative yield). j. i)nBu₄NF (100%); ii) TsOH/acetone (100%); iii) MsCl, pyr. (100%); iv) Na/acetone, reflux temp, overnight (99%).

conversion to an iodo derivative $(13)^9$ in quantitative yield, which possessed the same storeostructure as that of 1 and the required functional group for the coupling reaction with other segments.

Synthesis of segment **B**.

Treatment of the known triol (14) with dimethoxytoluene afforded a benzylidene derivative $(15)^{9,12}$ in 86% yield, which was converted to a silyl ether $(16)^9$ in four steps involving the hydroboration using 9-BBN. This compound (16) was submitted to Hanessian's bromination to open the acetal ring, and then the resulted benzoyl ester was transformed into an isobutyryl ester (17).⁹ Cyclization of 17 with LDA resulted in the desired δ -lactone $(18)^9$ in high yield. After partial reduction of 18 with DIBAL, the lactol so far obtained was reacted with 1,3-propanedithiol followed by protection of the OH groups to give an acyclic derivative $(19)^9$ in good overall yield. This compound would be coupled with the aforementioned 13 and the two silylated OH groups will be converted to the corresponding ester and ketonic CO groups, as seen in segment [B]. In particular, 19 may be used as a versatile intermediate for other aplysiatoxins.



a. PhCH(OMe)₂, TsOH/CH₂Cl₂ (86%). b. i) Swern oxi. (91%); ii) Ph₃P=CH₂THF, 0 $^{\circ}$ C \rightarrow room temp., (81%); iii) 9-BBN/THF, 0 $^{\circ}$ C \rightarrow room temp., overnight, then NaOH-H₂O₂ (93%); iv) TBDPSCI, Imd./DMF (100%). c. i) NBS, BaCO₃/CCl₄, reflux temp, 1 h (89%); ii) DIBAL-H/ PhMe, -78 $^{\circ}$ C, 20 min (88%); iii) 1 PrCOCI, pyr. (99%). d. LDA/THF, -78 \rightarrow 0 $^{\circ}$ C, 4 h (86%). e. i) DIBAL-H/PhMe, -78 $^{\circ}$ C, 30 min (95%); ii) 1,3-propanedithiol, BF₃OEt₂ (73%); iii) TBDMSCI, Imd./DMF (96%).

Synthesis of segment C.

According to the known procedure, 13 a thiol ester (22)⁹ could be synthesized from D-tartaric acid via an alcohol (20) and a cyano derivative (21).



a. i) TsCl, pyr.; ii) LiAlH4/THF; iii) aq. HCl-MeOH; iv) TsCl, pyr.; v) KCN/DMSO. b. i) HCl/MeOH; ii) aq. NaOH-THF; iii) tBuSH, DCC, DMAP/DMF

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 1.4, CHCl3j: H MMR (CDCl3) 6 1.01 (3H, d, J= 7 Hz), 1.30 (3H, s), 1.47 (3H, s), 2.27 (1H, m), 3.7 - 4.2 (4H, complex), 4.52 (1H, t, J= 4 Hz), 4.56 (2H, s), 5.76 (1H, d, J= 3 Hz), and 7.32 (5H, br.s). 5: Ciprag0435C1 [m/z 328.1435(M⁺)]: [alp²⁵ -6.2° (c), 7. CHCl3): IR (film) 3480 cm⁻¹; H MMR (CDCl3) 6 0.78 (3H, d, J= 7 Hz), 1.34 (3H, s), 1.39 (3H, s), 1.98 (1H, m), 3.5 - 4.3 (7H, complex), 4.56 (2H, s), and 7.32 (5H, br.s). 7: CigHp10a [m/z 277.1427 (M⁺-Me)]: [alp²⁵ -0.72° (c, 1.25, CHCl3): IH MMR (CDCl3) 6 0.97 (3H, d, J= 7.5 Hz), 2.90 (1H, d, J= 4, 6, 9. Hz), and 3.14 (1H, dt, J= 4.8, 9 Hz). as Ciprag0a [m/z 203.1751(M⁺-Me)]: [alp²⁵ -0.72° (c, 1.12, CHCl3): IR (film) 3520 cm⁻¹: 1H MMR (CDCl3) 6 0.75 (3H, d, J= 6.8 Hz), 0.91 (3H, d, J= 7.0 Hz), 1.37 (3H, s), 1.42 (3H, s), 1.77 (1H, m), 1.96(1H, m), 3.43 (1H, dd, J= 6, 9 Hz), 3.82 (1H, s), 4.10 (2H, m), 4.51 (1H, d, J= 12 Hz), 4.56 (1H, d, J= 7.9 Hz), 3.66 (1H, d, J= 7.9 Hz), 3.73 (1H, br.d, J= 9 Hz), 3.82 (1H, s), 4.10 (2H, m), 4.51 (1H, d, J= 12 Hz), 4.56 (2H, complex), 10: CajHa605Si [m/z 525.3042 (M⁺-Me)]; [alp²⁴ -10.0° (c 0.18, CHCl3); IR (film) 1680 and 1580 cm⁻¹: 1H MMR (CDCl3) 6 0.87 (3H, d, J= 7.3 Hz), 0.94 (3H, d, J= 6.3 Hz), 1.62 (1H, m), 1.70 - 1.88 (2H, complex), 1.94 (1H, m), 2.98 (2H, m), 3.59 (1H, dd, J= 3.74, 4Z, HZ), 11: Cag7Ha904I [m/z 552.3711 (M⁺-Me)]; [alp²⁴ -2.39° (c 1.02, CHCl3); IR (film) 1580 cm⁻¹: 1H MMR (CDCl3) 6 0.76 (3H, d, J= 7 Hz), 1.03 (3H, s), and 3.76 (1H, dd, J= 6.3 Hz), 1.33 (6H, s), and 3.18 (3H, s), 1.57 (3H, d, J= 7 Hz), 2.02 (1H, m), 2.29 (1H, dG, J= 3.18° (C 1.33; 1H MMR (CDCl3) 6 0.76 (3H, d, J= 7 Hz), 1.03 (3H, s), and 3.76 (1H, dd, J= 6.1 Hz), 1.33 (6H, s), and 3.
- 11. The purity could be determined based on the methyl signals due to TBDMS group in the reduction product [the desired S configuration: δ -0.021 and 0.017; the R configuration: δ -0.006 and 0.025].
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